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### Revisiting D'Adamo's Blood Type Diet: The Critical Role of Secreted Antigens in Digestive Health - An Evolutionary Perspective

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#### **ABSTRACT**

The theoretical construct of a 'Blood Specific Diet' advocated by Peter D'Adamo, on the premise that food lectins react differently with each blood type, has met with criticism from different quarters for different reasons. The approach followed in this paper is a paradigm shift from 'blood types' to 'secreted antigens', as the key factor, in the relationship between diet and disease (gastric / duodenal ulceration). The problem is analyzed from an evolutionary perspective, relying essentially on a built-in-set of biological mechanism in the form of secreted blood group substances, which confer a selective advantage in an environment of lectin-rich-diet. It is an evolutionary legacy that has been retained, to this day, in hunting-gathering societies. Lectins have been implicated as the trigger factor for peptic ulceration. Decreased susceptibility of secretors to peptic ulceration is closely related to the presence of glycoprotein form of blood group substances in large quantity In the brush border cells, which act as a defense against the mucous stripping effect of lectins in food. Due to their lectin binding affinity, secreted antigens also prevent the lectins from attaching to and damaging the cell membrane lining of the gut. Three primary factors namely, quantum of lectin intake, secretor status along with ABO phenotype and preferential affinity of H. pylori key-adhesins for competing H type 2 and Lewis-a and Lewis-b antigenic receptors, seem to play a pivotal role in determining the bacterial load, inflammatory response and severity of H. pylori infection leading to gastric / duodenal ulceration. Propensity for gastric ulceration is a bane of modern societies which are not so well adapted to withstand the onslaught of lectin-rich-diet.

#### **HYPOTHESIS**

The ability to secrete A-B-H substances in saliva and other glandular secretions was, perhaps, of paramount adaptive significance at a stage in the early history of man when human ancestors subsisted on raw food like other animals. Dietary habits shifted profoundly with the invention of fire making techniques, which greatly increased consumption of cooked food, The cultural revolution in food habits of early man resulted in relaxation of selection pressure on the dominant secretor gene (Se) necessary for survival in a

lectin-rich dietary environment. With the abatement of selection pressure, conditions were set for the recessive mutant gene 'se', to accumulate over a long period of time, leading to the establishment of ABH polymorphism in man. Evidence lending support to 'selection-relaxation' hypothesis is derived from the distribution of secretor types (which is purportedly related to history of dependence on raw food) in man and non-human primates. Serological investigations carried out on hundreds of apes and monkeys reveal that they are invariably secretors. The non-

human primates have a diversified diet, but they are primarily vegetarian and feed on ripe fruits, leaves, blades, and a plethora of wild plants available in their forage area. Their diet is most stressful in terms of its un-tempered lectin activity, which explains the absence of nonsecretors in monkeys and anthropoid apes. The hunter-gatherers obtain a bulk of their food from hunting, fishing and gathering insects and wild plants. Both in size and level of their economy, the hunting-gathering societies present the closest approximation, one can find, to the conditions under which early ancestors of man lived. The content of raw food is high in their diet. A very low incidence, bordering on virtual absence, of non-secretors (zero to 3%) in hunter-gatherers can be attributed to a surfeit of dietary lectins present in the food consumed by them. Population surveys across the world show a frequent occurrence of non-secretors (over 20%) in modern human societies. A significant increase in the incidence of non-secretors in modern human societies is, in all likelihood, the consequence of subdued lectin activity in cooked food (lectins being sensitive to heat), which forms a major component of their daily diet. The selection-relaxation hypothesis perceives the observed differences in incidence of non-secretors in man and non-human adaptive adjustment primates an environmental stress exerted by lectins in food on gastric mucosa. A piquant situation arises, more so in developed countries, due to abundance of food and a tendency among the affluents to consume lectin-rich food in excess, which makes them prone not only to ulceration but also to diseases of affluence viz., obesity, diabetes type 2 and cardiovascular diseases.

The concept of 'Blood Type Diet' advocated by Dr. Peter J. D'Adamo [1] is a contentious issue that has met with criticisms from different quarters for many different reasons [2,3,4,5]. Central to D'Adamo's theory of Blood Type Diet is his assertion that lectins in food react differently with each ABO blood type. According to him, lectins which interact with different blood types are

'incompatible and harmful', and that selection of different foods for A, B, AB and O blood types is important to minimize reactions with the lectins. D'Adamo's claim that there are many ABO specific lectins is not substantiated by findings of biochemical research. Investigations carried out on a wide variety of foods show that, "Most lectins in plant species are not ABO blood type specific. Even fewer Edible plants have lectins that are ABO type specific" [6]. Lectins found in commonly consumed foods are nonspecific in their reactivity with red cells of different blood types <sup>[6,7]</sup>. The theoretical construct of a blood group specific diet, on the ground that food lectins react differently with each blood type, therefore, remains unsubstantiated.

Addressing to the role of secretor factor, D'Adamo [1] writes on page 20 of 'Eat Right 4 your Type' "at this point you might be wondering about other blood type identifiers, such as positive / negative, or secretor / non-secretor ... These variations or subgroups within blood types play relatively insignificant roles".

Contrary to D'Adamo's assertion, there is ample evidence available in the published literature to support the critical importance of secreted ABH antigens, due to their ability to inhibit both specific as well as non-specific lectins [8,9,10].

Early indications of a protective role of secreted antigens appeared soon after Clarke et al., [11,12] reported that duodenal ulceration is likely to develop 50% more often in non-secretors of A-B-H substances than in secretors. Taking the likelihood of group least susceptible to duodenal ulcers (Secretors not O) as 1, the relative liability of O secretors was estimated as 1.35, of A and AB non-secretors as 1.6 and of O non-secretors as 2.5. It can be deduced from these statistics that O nonsecretors are two and a half times more likely to have duodenal ulcers compared with A, B and AB secretors. These early findings have been largely upheld by subsequent researches. While some of the data are open to criticism, a large body of observations made in hundreds of studies from many different countries show close association

between non-secretion and both duodenal and gastric ulceration [13,14,15]. Researches, in more recent years, have linked the incidence of ulcers to the presence of Helicobacter pylori, a Gramnegative bacterium [16,17,18,19], which colonizes the stomach and induces chronic gastritis, a long lasting inflammation of the stomach. About 10-20% of those colonized by H. pylori will ultimately develop gastric and duodenal ulcers [19]. Chronic gastritis can harm the stomach and duodenal lining by several mechanisms. One such mechanism involves the adherence of H. pylori to the epithelial cells by producing adhesins which bind to lipids and carbohydrates in the epithelial cell membrane <sup>[20]</sup>. In order to survive in the harsh acidic environment of the stomach, H. Pylori uses chemotaxis to avoid areas of low pH. It burrows into the mucus lining of the stomach to reach the epithelial cells beneath, where the pH is more neutral [21,22]. Colonization with H. pylori is not a disease in and of itself but a condition associated with a number of disorders of the upper gastrointestinal tract [19]. While both secretors as well as non-secretors may harbor H. pylori in their digestive tract, the inflammatory response, which is mediated in whole or in part, by host expression of secreted A-B-H and Lewis antigens in gastric mucosa, and the consequent severity of infection leading to duodenal / gastric ulceration, is more likely to occur in Group-O non-secretors than in A, B and AB secretors. Consistent with these observations, Mentis et al., [23] found persistent infection by H. pylori in 80% of non-secretors compared with 37% of secretors following attempted eradication therapy.

Corroborative evidence lending support to a protective role of secreted A-B-H substances against lectins in diet has also been derived from comparative study of secretor types in man and nonhuman primates <sup>[24]</sup>. Among hundreds of apes and monkeys tested by Wiener, Moor-Jankowski and coworkers <sup>[25]</sup>, all were found to be secretors. In striking contrast, the human population groups show wide variations in the incidence of non-secretors. The advanced (modern) human

societies, by and large, exhibit a high incidence of non-secretors (over 20%). The primitive human societies (Eskimo, Australian aborigines, Bushmen of South Africa, Natives of New Guinea, Onges of Andaman Islands and a number of North and South American tribes), on the other hand, show strikingly low incidence of non-secretors, diminishing to zero in some of the samples drawn from these human isolates [26]. Both in size and level of their economy, the hunting-gathering societies present the closest approximation, one can find, to the conditions under which early ancestors of man lived. A complete absence of non-secretors in non-human primates, exceedingly low incidence in hunting-gathering societies and a sustained high frequency in advanced human societies, clearly indicate that non-secretors are at a selective disadvantage in populations exposed to an environment of lectinrich-diet. (24). The ability to secrete A-B-H substances in saliva and other glandular secretions was, perhaps, of paramount adaptive significance at a stage, in the early history of man, when human ancestors subsisted on raw food, like other animals (24). With the invention of fire making techniques, food habits of early man witnessed a radical change from raw to cooked food. The cultural revolution in food habits, which greatly increased the consumption of cooked food, resulted in a reversal of negative selection pressure bearing upon non-secretor alleles (se), leading to it's establishment and maintenance in human populations, at frequencies much above the level of it's mutation rate [24].

Humans have lived by hunting and gathering for more than 99% of our evolutionary history [27]. The hunting-gathering economy, (which is essentially non-agrarian in nature), represents the original mean of subsistence common to all prehistoric humans during the old Stone Age [27]. Agriculture was introduced by some populations about 10,000 years ago, and their descendants possibly have some genetic adaptation to an agrarian diet [28,29]. However, many of the human populations shifted to an agrarian (cereal based)

diet more recently (< 100 generations ago) which, from an evolutionary point of view, is very short time for any measurable signs of adaptation [30]. Thus, when examining human diet from an evolutionary perspective, it makes sense that humans with an evolutionary novel agrarian diet consisting of cereals (wheat, rice, maze etc.), could suffer from diseases of affluence (obesity, cardiovascular diseases and diabetes type), due to insufficient adaptation [31]. The cereal based diet specific to agrarian societies could be an important environmental factor to initiate these diseases through lectins as cereal constituents with sufficient properties to cause leptin resistance [32]. Many metabolic factors are important in the onset and development of diseases of affluence. Considering the close connect between ABH secretion and dietary lectins, it will be of interest to study the role of secretor status as a susceptibility factor to leptin resistance and predisposition to diseases of affluence.

The example of ABH secretion does serve to explain how modern civilization has led to relaxation of selection pressure on secretor gene (Se) necessary for survival in lectin-richenvironment. However, there still remains the to understand biochemical basis quantitative and qualitative variability in blood group active substances in saliva and other glandular secretions, and their interaction with lectins in food. Lectins differ widely in their hemagglutinating reactions and in the susceptibility of these reactions to inhibition by saccharides. Most lectins agglutinate human erythrocytes of all groups and are usually referred to as nonspecific lectins or panagglutinins [33]. The group specific lectins, on the other hand agglutinate preferentially the human erythrocytes of a given blood type and form precipitates with corresponding soluble blood group substance [33]. It has been shown that among the large group of so called nonspecific lectins, there are many which react selectively with human and animal red cells [34], indicating that the nonspecific lectins too have a measure of specificity. The interaction of nonspecific lectins

with cells can, in many cases, be inhibited specifically by human saliva [8,35,36] and simple sugars [33,37,38,39]. It is clear from the results of hemagglutination and inhibition studies that the nonspecific lectins differ markedly in their reactivity and sugar binding specificity. While some of them are highly specific in their binding affinity for saccharides, others fail to be inhibited by any of the sugars or other chemical compounds. The sugar specificity of nonspecific lectins is not necessarily related to that of blood group determinants. Even so, some of these could be converted into blood group specific agglutinins in the presence of inhibiting sugar(s) [8,33,40,41]. Lectins often do not occur singly but exist as groups of closely related proteins or isolectins. Identification of sugar specificity of lectins in food and their inhibition reaction with human saliva can be expected to provide useful insight into the clinical significance of differences between secretors and non-secretors and their adaptive response to agrarian and non-agrarian foods.

An important issue that needs to be addressed is the role of dietary lectins in the pathogenesis of disease. Humans consume a significant amount of lectins as part of their daily diet. Although many of the dietary lectins are inactivated by proper heat treatment, some lectins found in wheat bran, wheat flour, wheat germ, peanuts, dry cereals, carrot, maize, apple, pumpkin, banana etc., survive cooking [7,39,42]. Grant and coworkers [43] demonstrated that the kidney beens that have been heated for several hours in slow cookers are likely to retain enough lectins to cause gastroenteritis, especially, if not pre soaked before cooking. Having survived many lectins are destroyed by digestion. However, enough remain to cause enteric signs and symptoms in man and animals. They bind to the surface epithelium of the digestive tract and lead to anti-nutritional, mild allergic or other subclinical effects in humans and animals [44,45]. The nutritional toxicity of the lectins is dependent on whether significant amount of lectins are systemically absorbed or not

[46]. Lectins which are not bound by the mucosa usually induce little or no harmful effects [47]. Lectins have been implicated as the causative several diseases. White consumed excessively by humans contains a high proportion of gluten and has agglutinating activity suggestive of lectins [48,49]. Peptides behaving in a lectin-like manner have been obtained form cleavage of gliadin in gluten [50] which bind to human intestinal mucosa and cause coeliac disease in people sensitive to gliadin in diet [51]. Sour dough lactic acid bacteria hydrolysegliadin peptides and inhibit their lectin like behaviour [52] which, perhaps, explains some of the unexplained health effects of probiotics [53]. Some other suspect lectin diseases triggered by foods are insulin dependent diabetes (trigger factor: tomato lectin and probably also wheat potato and peanuts) [54,55], rheumatoid arthritis (trigger factor: wheat lectin) [56], coeliac disease (trigger factor: wheat gliadin) [57], IgA nephropathy (trigger factor wheat lectin) [58], food poisoning from raw and under cooked kidney beans [59,60] etc.

Of particular interest is the implication of lectins for peptic ulcer disease. There is mounting evidence indicating that peptic ulceration is a lectin induced disease [61]. The toxicity of lectins has been linked with consumption of food with high lectin content. Foods such as beans, cereal grains, potatoes, nuts etc., which contain lectins in high concentration, if consumed in excess can interact with mucosa to cause acute gastrointestinal symptoms in experimental animals [39,51,62]. One of the effects observed in the small intestine of lectin-fed rodents is stripping away of mucous coat to expose naked mucosa, and overgrowth of the mucosa by abnormal bacteria and Protozoa [63]. Lectins also cause discharge of histamine from gastric mast cells, which stimulate acid secretion <sup>[64]</sup>. Freed <sup>[61]</sup> sums up these observations to conclude that, "the three main pathogenic factors for peptic ulcer - acid stimulation, failure of the mucous defense and abnormal bacterial proliferation (Helicobacter pylori) theoretically linked to lectins. If true, blocking of these effects by oligosaccharides would represent an attractive and more physiological treatment for peptic ulcers than suppressing stomach acid". An obvious question that arises is that if we all eat lectins as a part of our daily diet, why only some people develop ulcers and not others. One plausible explanation would be the presence of glycoprotein form of blood group substances in large amounts in brush border cells of the secretors which, by virtue of their binding affinity for lectins, mitigate their mucus stripping effect and protect the epithelium from heavy colonization of infecting bacteria.

The role of Lewis-b antigen, which biochemically related to ABO blood groups and secretor factor, and has been implicated as a putative receptor for Helicobacter pylori in the gastric mucosa, is debatable. Several studies have reported an increase of Lewis-b phenotype in H. pylori infected patients [65,66]. It has been suggested that H. pylori bacteria adhere to the epithelial cells by producing adhesins such as BabA, which bind to the Lewis-b antigen displayed on the surface of stomach epithelial cells [20]. Another adhesin, SabA binds to sialyl Lewis-x antigen expressed on gastric mucosa [67]. Clyne and Drumm [68], using flow cytometry to investigate the binding of H. pylori to Kato III cells and the primary gastric epithelial cells, found that adherence of the bacterium occurred in a manner independent of Lewis-a and Lewis-b expression. Alkout and coworkers [69] identified an H. pylori adhesin 61-kDa that binds H type 2, Lewis-a and Lewis-b antigens. They argued that if a key adhesin of H. pylori binds to H type 2 of epithelial cells, the presence of Lewis-b in the mucus of secretors will compete with greater effect for the 61-kDa adhesin and reduce colonisation of H. pylori. Alternatively, nonsecretors expressing Lewis-a in their gastric mucosa will compete less effectively with the adhesin and, as a result, will be more densely colonised by the bacterium. Probing relationship between host Lewis and ABO blood group phenotypes and prevalence of H. pylori

infection, Heneghan et al., [70] reported that, "although no in vivo relationship exists between H. pylori and preferential adhesion to the putative Lewis-b receptor, bacterial colonisation and ensuing inflammatory response may be influenced at least in part by host expression of ABO and Lewis-a blood group antigens". They found greater bacterial density among patients whose red cells expressed Lewis-a antigen (non-secretor phenotype). Consistent with these results, Heneghan et al., [68] observed that both acute and chronic inflammatory cells were present in greater quantity in the antral mucosa of non-secretors and that there was a positive correlation between density of H. pylori infection and the degree of lymphocyte and neutrophil infiltration in both secretors and non-secretors.

#### Conclusion

Understanding the clinical significance of ABH secretor status, Lewis subtypes and lectins in diet affords a valuable insight into the pathogenesis of peptic ulceration. It is hypothesized that ability to secrete A,B and H substances in saliva, mucus and other glandular secretions was, perhaps, of paramount adaptive significance at a stage in the early history of man when human ancestors subsisted on raw food [24]. Dietary lectins exert a strong selection pressure favoring secretors. Evolutionary trends in the establishment of ABH polymorphism strongly suggest an inverse relationship between the incidence of secretors and the component of raw food rich-inlectins in diet. The non-human primates who feed mostly on wild plants (fruits, leaves, tubers etc.), and consume untempered lectins in bulk, are invariably secretors [25]. The hunter-gatherers, with a high content of lectin-rich raw food in their diet, likewise, are mostly secretors. The nonsecretors are rare (zero to 3%) in these human isolates [26]. The modern human societies, on the other hand, show a sustained high frequency of non-secretors (over 20%) [26]. As many of the dietary lectins are heat sensitive, they get inactivated, partly or fully, in the cooked food

which forms a major component of their daily diet. A frequent occurrence of non-secretors in modern human societies is perceived as the consequence of relaxation of selection pressure on the secretor gene (Se) which has a selective advantage in a lectin-rich dietary environment [24]. Plants are a rich source of lectins. Foods with high lectin content (legume seeds, cereal grains, nuts etc.), if consumed in excess, in uncooked or partially cooked form, can cause gastrointestinal distress [39,51,62]. There is mounting evidence implicating lectins as the trigger factor for peptic ulceration<sup>[61]</sup>. Decreased susceptibility of secretors to ulceration can be attributed to the presence of water soluble blood group substances (glycoproteins) in large amount in brush border cells lining the gut, which act as a defense against the mucus stripping effect of lectins in food, and protect the epithelium from heavy colonization of Helicobacter pylori. Some other factors influencing the onset and development of peptic ulcer disease are, a lectin rich diet [24,39,51,62], ABO phenotype [11,12] and preferential affinity of H. pylori key-adhesins for competing H type 2, Lewis-a and Lewis-b antigenic receptors in gastric mucosa [69,70]. Proactive role of some seemingly unrelated factors such as leptin resistance, trypsin inhibitor types, passage into blood plasma of intestinal alkaline phosphatase, PTC tasting ability etc., which have a bearing on the etiology of peptic ulcers also need to be investigated. Propensity for ulceration is a bane of modern human societies which are not so well adapted to withstand the onslaught of a lectin-rich-diet (raw, unprocessed and partly cooked foods).

#### REFERENCES

- 1. D'Adamo PJ: Eat right 4 your type. Putnam, New York 1997.
- Klaper M: Blood Type Diet: Fact or Fiction. Toronto Vegetarian Association, Nov. 11, 2005. Website: www.earthsave.org.
- 3. Leila Cusack, Emmy De Buck, Veerie Compernolle, PhillippeVanderkerchkove:

- Blood type diets lack supporting evidence: A systematic review. The American Journal of Clinical Nutrition, 98(1): 99-104, 2013.
- Wang Jingzhou, Bibiana Garcia-Bailo, Daiva E. Nielsen, Ahmed El-Sohemy: ABO Genotype, Blood Type Diet and Cardiometabolic Risk Factors. 15 Jan. 2014, PLoS ONE 9(1): e84749 doi: 10.1371/journal. pone. 0084749. PMC 3893150.
- 5. Mary-Jean King "2: ABO Polymorphism and their putative biological relationship with disease". Human Blood Cells (Consequences Genetic of Polymorphisms and Variations). (2000-07-04), World Scientific Pub Co Inc. pp 44. ISBN 978-1860941962.
- Owen Foundation website: www.owenfoundation.com,
   Lectins in Edible Foods and ABO Reactions: An alphabetical list of Foods containing Lectins in Edible Plants and Animal sources. Oct. 2013.
- 7. Nachbar MS, Oppenheim JD: Lectins in the United States diet: a survey of lectins in commonly consumed foods and a review of the literature. Am. J. Clin. Nut. 33:2338-2345, 1980.
- 8. Bhatia HM, Boyd WC: Inhibition reactions of Fourteen "Nonspecific" Seed Extracts. Transfusion 2(2): 106-109, 1962.
- 9. Boyd WC: Lectins. Annals of New York Academy of Sciences. 169 (Article 1), pp. 168-190, 1970.
- 10. Grundbacher FJ: Lectins in precipitin reactions with soluble H substance of Human Saliva and Serum. Science 1973.
- 11. Clarke CA, Edwards JW, Haddock DRW, Howel-Evans AW, McConnell RB, Sheppard PM: ABO blood groups and secretor character in duodenal ulcer. Brit. Med. J. ii, 725-731, 1956.

- 12. 6. Clarke CA, Evans DA, price A, McConnell RB, Sheppard PM: Secretion of blood group antigens and peptic ulcer. Brit. Med. J. i, 603-7, 1959.
- 13. Prokop O, Uhlenbruck G: Human Blood and Serum Groups. New York, NY Wiley Interscience; pp. 390-722, 1969.
- 14. Mourant AE, Kopec AC, Domaniewska-Sobczak K: Blood Groups and Disease. London, Oxford University Press, 1978.
- 15. Garratty G: Do blood groups have a biological role? In, Immunology of Transfusion Medicine. ed. Garratty G. New York, NY Dekker; pp. 201-255, 1994.
- 16. Marshal BJ, Warren JR: Unidentified curved bacilli in the stomach of patients with gastritis and peptic ulceration. Lancet 1 (8390): 1311-5, 1984.
- 17. NIH Consensus Statement Online: Helicobacter pylori in peptic ulcer disease. Jan. 7-9, 1994, pp. 1-23.
- 18. Shistani A, Graham DY: Pathogenesis and therapy of gastric and duodenal ulcer disease. Medi. Clin. North-Am. 86(6): 1447-66, 2002.
- 19. Kusters JG, van Vliet AH, Kuipers EJ: Pathogenesis of Helicobacter pylori Infection. Clin. Microbiol. Rev. 19(3): 449-90, 2006.
- 20. Liver D, Arnqvist A, Ogren J, Frick IM, Kersulyte D, Incecik ET, Berg DE, Convucci A, Engstrand L, Boren T: Helicobacter pylori adhesin binding fucosylatedhisto-blood group antigen revealed by retagging. Science, 279(5349): 373-7, 1998.
- 21. Schreiber S, Konradt M, Groll C, Scheid P, Hanauer G, Werling HO, Josenhans C, Suerbaum S: The spatial orientation of Helicobacter pylori in the gastric mucus. Proc. Natl. Acad. Sci. U.S.A. 101(14): 5024-9, 2004.
- 22. Peterson AM, Krogfelt KA: Helicobacter pylori an invading organism? A review.

- FEMS Immunol. Med. Microbiol. 36(3): 117-26, 2003.
- 23. Mentis A, Blackwell CC, Weir DM, Spiladic C, Dialianas A, Skandalis N: ABO blood group, secretor status and detection of Helicobacter pylori among patients with gastric or duodenal ulcers. Epidemiol. Infect. 106, 221-229, 1991.
- 24. Bhalla V: ABH Polymorphism and Lectins in Diet: A Selection-Relaxation Hypothesis. In, Lectins-Biology, Biochemistry, Cinical Biochemistry, Vol. 7, Sigma Chemical Company, St. Louis, Missouri USA, PP. 299-303, 1990.
- 25. Wiener AS, Moor-Jankowski J: Blood groups of non-human primates and their relationship to the blood groups of man. In, Monkeys, Apes and Man. ed. Chiarelli AB. Academic Press; London; pp. 71-95, 1971.
- 26. Mourant AE, Kopec AC, Domaniewska-Sobczak K: Blood Groups and Disease. London, Oxford University Press, 1978.
- 27. Kuhn SL, Stiner MC: The antiquity of Hunter-gatherers. In, Hunter-gatherers: an interdisciplinary perspective. ed. Panter Brick C, Rowley Conwy P and Layton R. Cambridge, Cambridge University Press, 2001: pp. 99-142. [The Biosocial Society symposium series].
- 28. Coming CC, Shanahan F: Why is celiac disease so common in Ireland?Perspect. Biol. Med. 44: 342-52, 2001. [PubMed Abstract].
- 29. Rostani K, Malekzadeh R, Shahbazkhani B, Akbari MR, Catassi C: Coeliac disease in Middle Eastern countries: a challenge for the evolutionary history of this complex disorder? Dig. Liver Disease. 36: 694-7. [PubMed Abstract].
- 30. Temple NJ, Burkitt DP: Western diseases: Their dietary prevention and reversibility, Totowa, N.J., Humana Press; xiii, 453, 1994.

- 31. Freeman S, Herron JC: Evolution analysis, 3rd ed. Upper Saddle River; xiv, 802, 2004.
- 32. Jonsson T, Olsson S, Ahren B, Bog-Hansen TC, Dole A, Lindeberg S: Agrarian diet and diseases of affluence -Do evolutionary novel dietary lectins cause leptin resistance? BMC Endocrine Disorders. 5:10, 2005.
- 33. Sharon N, Lis H. Lectins: Cellagglutinating and sugar-specific proteins. Science, 177(4053): 949-59, 1972.
- 34. Bhalla V, Gaur V, Bhatia K: Lectin studies III. A survey of phytohaemagglutinins: Interaction of lectins with erythrocytes of ten vertebrate species, Vox Sanguinis, 35: 241-247, 1978.
- 35. Gibbons JR, Dankers I: Lectin-like constituents of food which react with components of Serum, Saliva and Streptococcus mutans. Appl. Environ. Microbiol. 41(4): 880-888, 1981.
- 36. Bhalla V, Bhatia K, Gaur V, Satsangi S: Specific differences in the inhibition reactions of seven anti-H type lectins. ActaAnthropogenetica, 1(2): 9-14, 1977.
- 37. Boyd WC: The specificity of the nonspecific. J. Immunology, 85(3): 2 21-29, 1960
- 38. Sachdeva Rosy, Bhalla V: Inhibition reactions of ten nonspecific seed extracts and their conversion into group specific lectins. J. Hum. Ecology, 8(1): 39-42, 1997.
- 39. Liener IE: Phytohemagglutinis: Their nutritional significance. J. Agric. Food Chem. 22: 17, 1974.
- 40. Goldstein IJ, Poretz RD: Isolation, physicochemical characterization and carbohydrate-binding specificity of lectins. In, The Lectins. Ed. Liener IE, Sharon N, Goldstein IJ. Academic Press Inc., Harcourt Bruce Jovanovich, Publishers, London, 1986.

- 41. Bhalla, V: Lectins as diagnostic reagents: Application to problems in Sero-Anthropology, and detection of new sources in Indian Flora. In, Contributions to Holistic Traditions in Anthropology in India. Ed. R.K. Bhattacharya, Anthropological Survey of India, Calcutta, 2000.
- 42. Concon JM, Newberg DS, Eades SN: Lectins in Wheat Gluten Proteins. J. Agric. Food Chem. 31: 939-941, 1983.
- 43. Grant G, More LJ, McKenzie NH, Pusztai A: The effect of heating on the haemagglutinaing activity and nutritional properties of bean (Phaseolusvulgaris) seeds. J. Sci. Food. Agric. 33: 1324-1326, 1982.
- 44. Van Damme JME, Peumans WJ, Pusztai A, Bardocz S: Handbook of plant lectins: Properties and Biomedical Applications. Chichester, John Wiley, ill. xiv. 1st ed. 1998.
- 45. Sharon N, Lis H: Lectins. Kluwer Academic Publishers. Dordecht; London. xviii, 2nd ed. 2003.
- 46. Pusztai A: In, Toxicants of plant origin III. Proteins and Amino Acids. Cheeks, PR; ed., CRC Press, Inc. Boca Raton. Florida.1987.
- 47. Pusztai A: Characteristics and consequences of interaction of lectins with intestinal mucosa (abstract). Arch. Latinoam Nutr. 44 (4 Suppl 1): 10s-15s, 1996.
- 48. Shewry PR, Halford NG: Cereal seed storage proteins: structures, properties and role in grain utilisation. J. Exp. Bot., 53: 947-58, 2002. [PubMed Abstract).
- 49. Freed DLJ: Lectins in Food: Their importance in health and disease. Journal of Nutritional Medicine, 2: 45-65, 1991.
- 50. Silane M, De Vincenzi M: Bio active antinutritional peptides derived from cereal protamines: A review. Nahrung 43: 175-84, 1999. [PubMed Abstract].
- 51. Freed DLJ, Buckley CH: Mucotractive effect of lectin. Lancet, i: 585-6, 1978.

- 52. De Cagno R, De Angelis M, Lavermicocca P, De Vincenzi M, Giovannini C, Faccia M. Gobbetti M: Proteolysis by sour dough lactic acid bacteria: effects on wheat flour protein fractions and gliadin peptides involved in human cereal intolerance. Appl. Environ. Microbiol. 68: 623-33, 2002. [PubMed Abstract].
- 53. Mercenier A, Pavan S, Pot B: Probiotics as biotherapeutic agents: Present knowledge and future prospects. Curr. Pharm. Des. 9: 175-91, 2003. [PubMed Abstract].
- 54. Uchigata Y, Spitalnik SL, Tachiwaki O, Salata KF, Notkins AL: Pancreatic islet cell surface glycoprotein containing Gal B(1-4)GNAc-R identified by cytotoxic monoclonal antibodies. J. Exp. Med. 165: 124-39, 1987.
- 55. Scott FW, Kolb H: Cow's milk and insulin dependent diabetes mellitus. Lancet, 348: 613, 1996.
- 56. Bond A, Kerr MA, Hay FC: Distinct oligosaccharides content of rheumatoid arthritis derived immune complexes. Arthr. Rheum. 38: 744-9, 1995.
- 57. Kolberg J, Sollid L: Lectin activity of gluten identified as wheat germ agglutinin. Biochem. Biophys. Res. Comm. 130: 867-72, 1985.
- 58. Coppo R, Amore A, Roccatello D: Dietary antigens and primary IgA nephropathy. J. Am. Soc. Nephrol. 2: 173-80, 1992.
- 59. Freed DLJ: Dietary Lectins and the anti-Nutritive Effects of Gut Allergy. Elsevier, Biomedical Press, North Holland. pp. 411-22, 1979.
- 60. Liener IE, Sharon N, Goldstein IJ: The Lectins: Properties, Functions and Application in Biology and Medicine. Academic Press, Orlando, FL; 1986.
- 61. Freed DLJ: Do dietary lectins cause disease? BMJ. 318: 1023-24, 1999.
- 62. Pusztai A, Clarke EMW, King TP: The nutritional toxicity of Phaseolusvulgaris lectins. Proc. Nutr. Soc. 38:115, 1978.

- 63. Banwell JG, Howard R, Kabir I, Costerton JW: Bacterial overgrowth by indigenous microflora in the PHA fed rat. Can. J. Microbiol. 34: 1009-13, 1988.
- 64. Greer F, Pusztai A: Toxicity of kidney bean (Phaseolusvulgaris) in rats: Changes in intestinal permeability. Digestion, 32: 42-6, 1985.
- 65. Martins LC, de Olivera Corvelo TC, [...] de Souza JT: ABH and Lewis antigen distribution in blood, saliva and mucous & H. pylori infection in gastric ulcer patients. World J. Gastroenterol. 12(7): 1120-4, 2006.
- 66. Sheu BS, Sheu SM, Yang HB, Huang AH, Wu JJ: Host gastric Lewis expression determines the bacterial density of Helicobacter pylori in babA2genopositive infection. Gut 52: 927-32, 2003.
- 67. Mahdavi J, Sonden B, Hurtig M, [...] Boren T: Helicobacter pylori SabA Adhesion in persistent infection and chronic inflammation. Science 297 (5581): 573-8, 2002.
- 68. Clyne M, Drumm B: Lewis blood group antigen expression and adherence of Helicobacter pylori to gastric cells. Gastroenterology 113: 72-80, 1997.
- 69. Alkout AM, Blackwell CC, Weir DM, Roxton IR, Elton RA, Luman W, Palmer K: Isolation of a cell surface component of Helicobacter pylori that binds H type 2, Lewis a and Lewis b antigens. Gastroenterology 112: 1179-87, 1997.
- 70. Heneghan MA, Moran AP, Feeley M, Egan EL, Goulding J, Connolly CE, McCarthy CF: Effects of host Lewis and ABO blood group antigen expression on Helicobactor pylori colonisation density and the consequent inflammatory response. Article first publishedonline 17 Jan. 2006. doi: 10.1111/j.1574-695X.1998.tb01135.x.