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Serum Calprotectin as a Non-invasive Diagnostic Marker for Spontaneous Bacterial Peritonitis in Egyptian Cirrhotic Patients

Authors

Amir Helmy¹, Ahmed El Saady Khayyal¹, Sara M Abdelhakam², Shereen A Saleh¹, Hesham Abouellail¹, Sameh Abdelraouf¹, Abdel Rahman Esam¹, Marwa Rushdy³

¹Department of Internal Medicine, Faculty of Medicine, Ain Shams University, Cairo, Egypt
²Department of Tropical Medicine, Faculty of Medicine, Ain Shams University, Cairo, Egypt
³Department of Clinical and Chemical Pathology, Faculty of Medicine, Ain Shams University, Cairo, Egypt
*Corresponding Author

Sara M Abdelhakam, MD

Assistant Professor of Tropical Medicine, Department of Tropical Medicine, Faculty of Medicine, Ain Shams University, Abbassia, Cairo, Egypt. Email: saratropical@yahoo.com, Telephone: (+2) 01001601548

ABSTRACT

BACKGROUND and AIM: Delayed diagnosis of spontaneous bacterial peritonitis (SBP) is associated with high mortality in cirrhotic patients. Therefore, there is a need for a rapid non-invasive diagnostic tool. This work aimed at evaluating the role of serum calprotectin as a non-invasive diagnostic marker for SBP in comparison to C-reactive protein (CRP).

METHODS: 75 cirrhotic patients were included and divided into three groups. **Group A:** 25 patients with cirrhotic ascites and SBP diagnosed by presence of PMNL $\geq 250/\text{mm}^3$ in ascitic fluid with or without positive ascitic fluid culture. **Group B:** 25 patients with cirrhotic ascites without SBP. **Group C:** 25 cirrhotic patients with no clinical or ultrasound evidence of ascites as a control group. All were subjected to complete clinical evaluation, routine laboratory investigations, serum calprotectin, CRP levels and diagnostic abdominal paracentesis (only for group A and B).

RESULTS: Group A showed significantly higher levels of ascitic fluid total leukocytic count, polymorphonuclear leukocytes and total protein (P<0.001) in comparison to Group B. Group A had significantly higher serum calprotectin and CRP levels than other groups. There were significant positive correlations between serum calprotectin and serum WBC, CRP and ascitic fluid TLC, PMNL and total protein among group A. Serum calprotectin and CRP had the most significant diagnostic performance in detection of SBP (AUC=0.976 and 1.000 respectively, P<0.001). A cut-off level of serum calprotectin \geq 46.0 μ g/mL showed a sensitivity of 100%, specificity 92% and diagnostic accuracy 96%; while that of CRP \geq 9.0 mg/dL showed a sensitivity of 100%, specificity 100% and diagnostic accuracy 100%.

CONCLUSION: Serum calprotectin in combination with CRP may be useful for non-invasive diagnosis of SBP.

KEY WORDS: Serum calprotectin, Spontaneous bacterial peritonitis, Liver cirrhosis, Ascites

INTRODUCTION

Spontaneous bacterial peritonitis (SBP) is defined as an infection of the ascitic fluid in the absence of visceral perforation or intra-abdominal inflammation such as abscess, acute pancreatitis or cholecystitis ^(1, 2). It is the most frequent infection in patients with liver cirrhosis and ascites with an incidence varying from 10%-30% ^(3, 4)

It has been proven that delayed diagnosis of SBP is associated with high mortality. Thus, an accurate biomarker for the early identification of SBP would be of great diagnostic value. Diagnosis of SBP is based on the presence of neutrophils more than 250/mm³ in ascitic fluid ⁽⁵⁾. Currently, differential cell count is usually performed by a manual method using light microscopy and counting chambers. However, the diagnosis is often delayed when laboratory personnel are not readily available or in the private practice setting where specimens are sent to an offsite laboratory. Alternative methods using polymorphonuclear leukocytes automated (PMNL) counting have been developed but unfortunately, their diagnostic accuracies are limited ⁽⁶⁾.

Also, the prolonged time of ascitic fluid culture limits its utility for directing antibiotic selection in acute care settings. The culture has also been reported to be negative in approximately 20% of patients with clinical manifestations suggestive of SBP and an ascitic PMNL count of >250/mm³, so-called culture-negative neutrocytic ascites (CNNA). On the other hand, a low ascitic PMNL count (<250/mm³) with positive culture can also occur in another SBP variant called monomicrobial non-neutrocytic bacterascites ⁽⁷⁾.

Calprotectin is an abundant, calcium- and zincbinding protein found mainly in neutrophils ⁽⁸⁾, and its presence in body fluids is proportional to the influx of neutrophils ⁽⁹⁾. It is an acute phase inflammatory reaction protein which exerts regulatory, antimicrobial and anti-proliferative functions. It can halt bacterial growth, playing an important role in non-specific immune reactions (10)

A recent study for measurement of ascitic fluid calprotectin by reagent strips as a bed side test for rapid diagnosis of SBP showed high specificity (83%) and sensitivity (100%) ⁽⁶⁾. Another study reported a significant correlation between elevated fecal calprotectin level and the occurrence of some complications in cirrhotic patients such as hepatic encephalopathy and SBP ⁽¹¹⁾. However, the role of serum calprotectin in diagnosing SBP remains unexplored.

This work aimed at evaluating the role of serum calprotectin as a non-invasive diagnostic marker for spontaneous bacterial peritonitis in Egyptian cirrhotic patients in comparison to C-reactive protein.

PATIENTS AND METHODS

This case control study was conducted on 75 cirrhotic patients admitted to Internal Medicine and Tropical Medicine Departments of Ain Shams University Hospital during the period from June 2015 to February 2016. The patients were divided into three groups. **Group A** included 25 patients with cirrhotic ascites and spontaneous bacterial peritonitis (SBP). They were diagnosed by presence of PMNL more than 250/mm³ in ascitic fluid with or without positive ascitic fluid culture. **Group B** included 25 patients with cirrhotic ascites without clinical or laboratory evidence of SBP and with ascitic fluid PMNL count less than 250 and negative culture. **Group C** included 25 cirrhotic patients with no clinical or ultrasound evidence of ascites as a control group.

Informed written consent was obtained from each patient prior to inclusion. The study protocol was approved by the Research Ethical Committee of Faculty of Medicine, Ain Shams University according to the ethical guidelines of the 1975 Declaration of Helsinki.

Patients with secondary causes of peritonitis such as T.B or malignancy; surgical causes of peritonitis such as intra-abdominal abscess, appendicitis, or pancreatitis; history of abdominal surgery in the previous 3 months; previous infection or antibiotic usage one month before presentation; hepatocellular carcinoma; portal or splenic vein thrombosis; inflammatory bowel disease; as well as those with diabetes mellitus were excluded.

All included patients were subjected to the following:

- 1. Complete history taking and thorough clinical examination.
- 2. Laboratory investigations:
 - a) Complete Blood Count (CBC): was done on Coulter LH 750 hematology analyzer (Beckman Coulter International SA, Rue Juste-Olivier 22, PO Box 1059, CH 1260 Nyon, Switzerland).
 - b) Liver profile: alanine aminotransferase (ALT), aspartate aminotransferase (AST), serum albumin, total and direct bilirubin, alkaline phosphatase, prothrombin time and INR.
 - c) Renal profile: creatinine and blood urea.

Liver and renal profiles were done on Synchron CX-5 Delta auto-analyzer using its commercially available reagents (Beckman Instruments Inc.; Scientific Instruments, Division, Fullerton, CA 92634, 3100, USA).

- d) Viral markers including: HBsAg, HBcAb IgM and IgG and HCV Ab by third generation ELISA.
- e) Serum C-reactive protein (CRP): was quantified by using the latex slide test (semi-quantitative test) according to the manufacturer's (Teco Diagnostics, Anaheim, CA 92807, U.S.A) instructions.
- f) Serum calprotectin level: Assay was carried out using a commercially available enzyme-linked immunosorbent assay (ELISA) kit supplied by (Shanghai Sunred Biological Technology Co) with normal assay

range: 0.15-40 µg/ml. The kit used a double-antibody sandwich enzymelinked immunosorbent assay (ELISA) assay the level of Human Calprotectin (CALB) in samples. Calprotectin (CALB) was added to monoclonal antibody Enzyme well which was pre-coated with Human Calprotectin monoclonal antibody, after incubation Calprotectin antibodies labeled with biotin, and combined with Streptavidin-HRP were added to form immune complex; after incubation Chromogen Solution A, B added, the color of the liquid changed into the blue, and at the effect of acid, the color finally became yellow. The chroma of color and the concentration of the Human Substance Calprotectin (CALB) of sample were positively correlated.

- 3. Abdominal ultrasonography: To assess liver size, liver echogenicity, amount of ascites, spleen size, and any focal lesions.
- 4. Diagnostic abdominal paracentesis (for patients in group A and B): It was performed under ultrasonographic guidance and aseptic condition. Three samples of ascitic fluid, each of ten centimeters, were taken from each patient at time of admission and sent immediately bacteriological, chemical for and pathological examination with assessment ascitic polymorphonuclear fluid leukocytic count (PMNL), total cell count, ascitic fluid albumin, total protein, lactate dehydrogenase (LDH), glucose and ascitic fluid culture. The sample for culture was inoculated in aerobic blood culture bottles (Egyptian Diagnostic Media, Cairo, Egypt) at the bedside, and then it was incubated for 3 successive days at 37°C with daily subculture on blood, MacConkey and chocolate agars.

5. Serum-ascites albumin gradient (SAAG) was estimated as (serum albumin - ascitic fluid albumin).

Statistical analysis

Collected data were coded, tabulated, and statistically analyzed using IBM SPSS statistics (Statistical Package for Social Sciences) software version 22. Descriptive statistics such as minimum and maximum of the range, as well as mean \pm SD, were used for quantitative parametric data. Median and inter-quartile ranges were used for quantitative non-parametric data, while number and percentage were used for qualitative data.

Inferential analyses were done for quantitative variables using independent t-test in cases of two independent groups with parametric data. Correlations were done using Pearson Correlation for numerical parametric data, and Spearman Rho test for numerical non- parametric as well as qualitative data. Receiver operating characteristics (ROC) curve was used to evaluate the performance of different tests, and DeLong test was used to compare between the areas under curves (AUCs).

Diagnostic characteristics were calculated as follows:

- Sensitivity = [true positive / (true positive+ false negative)] × 100
- Specificity = [true negative / (true negative+ false positive)] × 100
- Positive likelihood ratio (LR+) = (true positive / false positive) × 100
- Negative likelihood ratio (LR-) = (false negative / true negative) × 100
- Diagnostic accuracy = [(true positive + true negative) / total cases] × 100.
 - P value < 0.05 was considered statistically significant and P < 0.01 as highly significant.

RESULTS

The present study was conducted on 75 cirrhotic patients divided into three groups:

Group A included 25 cirrhotic patients with ascites and spontaneous bacterial peritonitis (SBP) at the time of presentation. They were 16 (64%) males and 9 (36%) females. Their mean age was 55.2 ± 6.9 years. Twenty three patients had HCV and two patients had HBV infection. Their main clinical presentations were abdominal pain in 22 patients (88%), fever in 21 (84%) and jaundice in 10 patients (40%). Among this group, 19 patients (76%) had PMNL >250/mm³ and positive ascitic fluid cultures (i.e. culture-positive SBP); the organisms isolated were Escherichia coli (12 patients), Citrobacter (three patients), Klebsiella (two patients), Acinetobacter (one patient) and Enterococci (one patient). The remaining six patients (24%) had PMNL >250/mm³ without positive bacteriological culture of ascitic fluid (i.e. culture-negative neutrocytic ascites, CNNA).

Group B included 25 cirrhotic patients with ascites with no clinical or laboratory evidence of SBP (sterile cirrhotic ascites). They were 17 (68%) males and 8 (32%) females. Their mean age was 54.0 ± 6.2 years. Twenty four patients had HCV and one patient had HBV infection.

Group C included 25 cirrhotic patients without ascites. They were 14 (56%) males and 11 (44%) females. Their mean age was 54.2 ± 7.3 . All patients of this group were positive for HCV.

There was no significant difference between the three studied groups regarding the age or gender with P value > 0.05.

Regarding the Child classification among the three studied groups, group A included 23 patients (92%) with Child score C and 2 patients (8%) with Child score B. Group B included 19 patients (76%) with Child score C and 6 patients (24%) with Child score B. All patients (100%) of group C had Child score A. There was statistically significant difference between groups A and C, and between groups B and C regarding the Child score with P value < 0.05.

Table (1) shows comparison between the three studied groups regarding demographic data and laboratory investigations. Group A had significantly high serum total leukocytic count and low platelet count (P<0.001).

Table (1): Comparison between the three studied groups regarding demographic data and laboratory investigations:

| Variable | | Group A (N=25) | Group B (N=25) | Group C (N=25) | A/B | A/C | B/C |
|-----------------------------|---------|-------------------|-------------------|-------------------|--------------------|--------------------|----------|
| Age (years) | Mean±SD | 55.2±6.9 | 54.0±6.2 | 54.2±7.3 | t=0.647 | t=0.497 | t=0.104 |
| | | | | | p=0.521 | p=0.621 | p=0.918 |
| Total bilirubin (0.2- | Mean±SD | 3.1±2.3 | 2.3±1.3 | 1.7±0.8 | t=1.513 | t=2.926 | t=2.000 |
| 1.2 mg/dL) | | | | | p=0.137 | p=0.005* | p=0.050* |
| ALT (7-40 IU/L) | Mean±SD | 46.5±19.6 | 50.7±22.7 | 49.0±20.0 | t=0.307 | t=0.888 | t=1.183 |
| | | | | | p=0.760 | p=0.379 | p=0.243 |
| AST (7-37 IU/L) | Mean±SD | 51.9±20.4 | 53.1±23.3 | 52.8±22.8 | t=0.460 | t=2.082 | t=1.424 |
| | | | | | p=0.648 | p=0.043* | p=0.161 |
| Serum Albumin | Mean±SD | 2.6±0.2 | 2.7±0.4 | 3.2±0.3 | t=0.638 | t=11.34 | t=11.29 |
| (3.5-5.3 g/dL) | | | | | p=0.526 | p<0.001* | p<0.001* |
| INR | Mean±SD | 1.6±0.3 | 1.5±0.5 | 1.2±0.1 | t=0.424 | t=8.518 | t=7.097 |
| | | | | | p=0.673 | p<0.001* | p<0.001* |
| HB (g/dL) | Mean±SD | 10.1±1.4 | 9.7±1.4 | 10.1±1.2 | t=1.073 | t=0.065 | t=1.063 |
| | | | | | p=0.289 | p=0.948 | p=0.293 |
| WBC (×10 ³ / μL) | Mean±SD | 15.3±3.5 | 6.8±2.7 | 6.5±2.0 | t=9.742 | t=10.99 | t=0.425 |
| | | | | | p=< 0.001 * | p=< 0.001 * | p=0.673 |
| PLT (×10 ³ / μL) | Mean±SD | 77.9±26.0 | 153.4±65.3 | 189.0±55.6 | t=5.370 | t=9.046 | t=2.076 |
| | | | | | p<0.001* | p<0.001* | p=0.043* |

^{*}Significant, ALT: Alanine aminotransferase, AST: Aspartate aminotransferase, INR: International normalization ratio, HB: Haemoglobin, WBC: White blood cells, PLT: Platelets.

Table (2) shows comparison between groups A and B regarding ascitic fluid analysis. Group A showed significantly high levels of ascitic fluid

total leukocytic count, polymorphonuclear leukocytes and total protein (P<0.001) in comparison to group B.

Table (2): Comparison between groups A & B regarding ascitic fluid analysis

| Variable (Mean±SD) | Group A (N=25) | Group B (N=25) | t | р |
|-----------------------|-------------------|-------------------|--------|---------|
| TLC (cell/mL) | 1307.5±712.3 | 8.3±2.2 | 9.120 | <0.001* |
| PMNL (cell/mL) | 551.3±71.8 | 6.3±1.4 | 37.932 | <0.001* |
| Albumin (g/dL) | 0.6±0.2 | 0.7±0.3 | 0.778 | 0.440 |
| Total Protein (g/dL) | 2.7±0.3 | 1.5±0.2 | 14.266 | <0.001* |
| SAAG | 1.9±0.5 | 2.0±0.6 | -0.707 | 0.483 |
| LDH (IU/L) | 139.2±73.1 | 138.6±78.5 | 0.030 | 0.976 |
| Glucose (mg/dL) | 105.1±13.6 | 116.1±31.7 | -1.598 | 0.117 |

*Significant

TLC: Total leukocytic count, PMNL: polymorphonuclear leukocytes, SAAG: serum-ascites albumin gradient, LDH: Lactate dehydrogenase.

Table (3) shows that group A had significantly higher serum calprotectin and C-reactive protein levels than other groups (P<0.001).

Table (3): Comparison between the three studied groups regarding serum calprotectin and CRP

| Variable | Group A | Group B | Group C | ^ | ^ | A |
|---------------------------------------|-----------|-----------|-----------|---------------------|---------------------|---------|
| | (N=25) | (N=25) | (N=25) | A/B | A/C | B/C |
| CRP (mg/dL) -Mean±SD -Range | 36.7±16.6 | 4.7±1.6 | 5.0±1.5 | t=9.593 | t=9.530 | t=0.551 |
| | 12.0–68.0 | 2.0–7.0 | 2.0-7.0 | p<0.001 * | p<0.001 * | p=0.584 |
| Calprotectin (μg /mL) -Mean±SD -Range | 61.6±7.3 | 24.8±14.8 | 24.4±11.8 | t=11.364 | t=13.431 | t=0.038 |
| | 47.9–73.7 | 8.9–46.7 | 6.6–39.0 | p<0.001 * | p<0.001 * | p=0.970 |

[^]Independent test, *Significant, CRP: C-reactive protein.

Table (4) and Figure (1) show correlation between serum calprotectin and different parameters among the studied groups. There were

significant positive correlations between serum calprotectin and serum WBC, CRP & ascitic fluid TLC, PMNL and total protein among group A.

Table (4): Correlation between serum calprotectin and different parameters among the studied groups.

| Variables | Group A (N=25) | | Group B (N=25) | | Group C (N=25) | |
|------------------|----------------|---------|----------------|-------|----------------|-------|
| | r^ | р | r^ | р | r^ | р |
| Age | -0.273 | 0.186 | 0.061 | 0.779 | -0.348 | 0.089 |
| Child score | 0.246 | 0.243 | 0.290 | 0.478 | 0.045 | 0.964 |
| Bilirubin | -0.026 | 0.900 | 0.183 | 0.391 | -0.127 | 0.545 |
| ALT | -0.112 | 0.594 | -0.045 | 0.833 | -0.137 | 0.512 |
| AST | -0.100 | 0.633 | 0.017 | 0.936 | -0.215 | 0.302 |
| Serum Albumin | 0.100 | 0.634 | -0.053 | 0.804 | 0.313 | 0.128 |
| INR | 0.140 | 0.504 | -0.011 | 0.961 | -0.351 | 0.085 |
| Hb | 0.109 | 0.604 | -0.101 | 0.638 | -0.274 | 0.185 |
| WBC | 0.969 | <0.001* | -0.023 | 0.915 | -0.197 | 0.344 |
| PLT | -0.010 | 0.961 | 0.141 | 0.512 | -0.207 | 0.321 |
| CRP | 0.963 | <0.001* | 0.011 | 0.960 | -0.039 | 0.855 |
| AF TLC | 0.816 | <0.001* | 0.180 | 0.399 | | |
| AF PMNL | 0.973 | <0.001* | -0.303 | 0.150 | | |
| AF Albumin | -0.047 | 0.824 | 0.321 | 0.126 | | |
| AF Total Protein | 0.519 | 0.029* | -0.219 | 0.294 | | |
| SAAG | 0.012 | 0.955 | -0.085 | 0.693 | | |
| AF LDH | 0.080 | 0.704 | -0.013 | 0.950 | | |
| AF Glucose | 0.050 | 0.812 | -0.126 | 0.559 | | |

[^]Spearman correlation, *Significant.

ALT: Alanine aminotransferase, AST: Aspartate aminotransferase, INR: International normalization ratio, HB: Haemoglobin, WBC: White blood cells, PLT: Platelets, CRP: C-reactive protein, AF: Ascitic fluid, TLC: Total leukocytic count, PMNL: polymorphonuclear leukocytes, SAAG: serum-ascites albumin gradient, LDH: Lactate dehydrogenase.

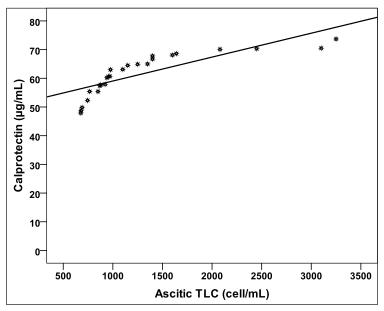


Figure (1): Correlation between serum calprotectin and ascitic fluid TLC among group A. There was a significant positive linear correlation ($r^{-}=0.816$, p<0.001).

Table (5) and Figure (2) show that serum calprotectin and CRP had the most significant diagnostic performance in detection of spontaneous bacterial peritonitis (SBP) (AUC=0.976 and 1.000 respectively, P<0.001).

On comparing the AUC of ROC curve of serum CRP and serum calprotectin in differentiating between group A (with SBP) and group B (with sterile cirrhotic ascites), it was found that P value was non-significant (P=0.242).

Table (5): Performance of serum WBC, CRP and calprotectin in detection of SBP.

| Variable | AUC | SE | P | 95% CI | | | | |
|--------------|--------------------------|---------|---------|-------------|--|--|--|--|
| Calprotectin | 0.976 | 0.012 | <0.001* | 0.500-1.000 | | | | |
| CRP | 1.000 | 0.000 | <0.001* | 1.000-1.000 | | | | |
| WBC | 0.625 | 0.080 | 0.130 | 0.500-0.782 | | | | |
| | Comparison between AUCs# | | | | | | | |
| | CRP | WBC | | | | | | |
| Calprotectin | 0.242 | <0.001* | | | | | | |
| CRP | | <0.001* | | | | | | |

SBP: spontaneous bacterial peritonitis, CRP: C-reactive protein, WBC: White blood cells, AUC: Area under curve, SE: Standard error, CI: Confidence interval, #DeLong test, *Significant.

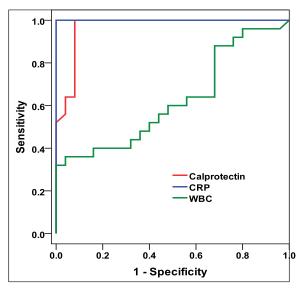


Figure (2): ROC curve showing the diagnostic performance of serum WBC, CRP and calprotectin in detection of SBP.

In differentiation between groups A & B, a cut-off level of serum calprotectin≥46.0 μg/mL showed a sensitivity of 100%, specificity 92% and diagnostic accuracy 96%; while that of CRP≥9.0

mg/dL showed a sensitivity of 100%, specificity 100% and diagnostic accuracy 100%. Thus, serum calprotectin≥ 46.0 and CRP≥9.0 were excellent parameters in detection of SBP (**Table 6**).

Table (6): Value of suggested cut-off levels of serum calprotectin and CRP in detection of SBP

| Character | Value | 95% CI | | | | | |
|--|--------|--------|--------|--|--|--|--|
| Calprotectin $\geq 46.0 \; (\mu g/mL)$ | | | | | | | |
| Sensitivity | 100.0% | 100.0% | 100.0% | | | | |
| Specificity | 92.0% | 90.0% | 92.5% | | | | |
| Positive likelihood ratio (LR+) | 12.5 | 52.4 | 800.4 | | | | |
| Negative likelihood ratio (LR-) | 0.0 | 0.0 | 0.0 | | | | |
| Diagnostic accuracy (DA) | 96.0% | 94.9% | 96.3% | | | | |
| CRP ≥ 9.0 (mg/dL) | • | | - | | | | |
| Sensitivity | 100.0% | 100.0% | 100.0% | | | | |
| Specificity | 100.0% | 100.0% | 100.0% | | | | |
| Positive likelihood ratio (LR+) | | | | | | | |
| Negative likelihood ratio (LR-) | 0.0 | 0.0 | 0.0 | | | | |
| Diagnostic accuracy (DA) | 100.0% | 100.0% | 100.0% | | | | |

⁻⁻ Cannot be calculated.

DISCUSSION

The diagnosis of SBP is still based on diagnostic paracentesis which is an invasive manoeuvre with some complications like ascitic fluid leakage from the point of needle insertion into the skin,

bleeding, and infection ⁽¹²⁾. In addition, the ascitic fluid culture takes a long time and may be negative in the entity of culture-negative neutrocytic ascites (CNNA) ⁽⁷⁾.

Therefore, there is a need for other non-invasive rapid diagnostic tools. Any serum biological marker that could strongly predict SBP may obviate the need for paracentesis while increasing the patient's chance of survival by expediting the diagnosis and treatment of SBP ⁽¹³⁾.

C-reactive protein (CRP) is an acute phase reactant which binds to different substrates. It activates complements, takes part in cytokine secretion, and increases the phagocytosis of leukocytes. CRP has been previously reported to be a reliable predictor of SBP and an index of improvement ⁽¹⁴⁾.

Calprotectin is an acute phase reactant secreted predominantly by the neutrophils and plays a vital role in infection and inflammation. It serves as a potential diagnostic marker for various inflammatory diseases ⁽¹⁵⁾. It has been previously reported that the measurement of fecal calprotectin concentration could serve as a screening tool for SBP ⁽¹¹⁾. However, the use of serum calprotectin in diagnosis of SBP remains unexplored.

This study aimed at evaluating the role of serum calprotectin as a non-invasive diagnostic marker for SBP in Egyptian cirrhotic patients in comparison to CRP.

Regarding the ascitic fluid cell count in the current study, there were higher levels of ascitic TLC and PMNL count in SBP group in comparison to the group with sterile cirrhotic ascites, and this result is in agreement with *Girón-González et al.* (16), *Yildirim et al.* (17) and *Abdel-Razik et al.* (18).

As regards ascitic fluid total protein, it was significantly higher in SBP group than in sterile cirrhotic ascites group (P<0.001). This result is in agreement with *Reginato et al.* ⁽¹⁹⁾.

Regarding the SAAG value, it was higher than 1.1 in SBP group with no statistically significant difference when compared to sterile cirrhotic ascites group, and this was in agreement with *Salama et al.* ⁽²⁰⁾. This finding can be explained by the study of *Desai et al.* ⁽²¹⁾, who denoted that regardless the severity of liver disease, those with lower ascitic albumin and higher SAAG levels are

less likely to have a satisfactory immune response with poor clearance of infection.

In this study, there was no significant difference between SBP group and the group with sterile cirrhotic ascites as regards the ascitic fluid glucose, and this is in agreement with *Bibi et al*. (22)

Regarding the relation between occurrence of SBP and Child-Pugh class, we found that 92% of patients with SBP were in Child C class compared to 76% of patients with sterile cirrhotic ascites, and this result is close to *Abdel-Razik et al.* (18) who found that about 75% of the patients who developed SBP were in Child C class, with the remainder were in class B.

In the current study, we found significantly higher levels of serum calprotectin in SBP patients than in the other two groups.

Ali et al. (23) found that ascitic calprotectin level showed significantly higher value in patients with SBP than those without SBP.

Lutz et al. (24) also reported that ascitic calprotectin could be used as a test for SBP, but test performance was further improved by calculating the ratio between ascitic calprotectin and ascitic total protein. They also reported that this ratio could provide prognostic information on short-term survival of SBP patients.

Gundling et al. (11) found also that fecal calprotectin levels were elevated significantly in cirrhotic patients with SBP in comparison to those without SBP.

In our study, we found a significant positive correlation between higher serum calprotectin level and elevated white blood cell count among SBP patients. This is consistent with *Abdel-Razik*, *et al.* ⁽²⁵⁾. This finding can be explained by the fact that serum calprotectin is an acute phase inflammatory reaction protein exerting regulatory, antimicrobial and anti-proliferative functions ⁽¹⁰⁾. In this study, we found that high serum calprotectin level was significantly correlated with high ascitic fluid TLC and PMNL levels in SBP group. Also, *Burri et al.* ⁽⁶⁾, *Abdel-Razik et al.* ⁽¹⁸⁾, *and Ali et al.* ⁽²³⁾, found that ascitic calprotectin

levels were correlated well and reliably with ascitic PMNL counts, and the samples with PMNL > $250/\mu L$ also had higher ascitic calprotectin levels than the samples with PMNL $\leq 250/\mu L$ in their studies.

The correlation between high serum & ascitic calprotectin with high ascitic TLC and PMNL levels can be explained by the study of *Soyfoo et al.* ⁽⁹⁾, who denoted that calprotectin presence in body fluids is proportional to the influx of neutrophils.

In our study, there was a significant positive correlation between serum calprotectin and CRP, and this result is in agreement with $Rizk\ et\ al.$ (26). We found also that both serum calprotectin and CRP were significantly higher in group A (with SBP) than group B (with sterile cirrhotic ascites) (P < 0.001).

In the current study, a cut off level of ≥ 9.0 (mg/dL) for serum CRP in detecting SBP showed a highly significant diagnostic performance with AUC 1.000 (P <0.001), sensitivity 100% and specificity 100%. *Rizk et al.* ⁽²⁶⁾ reported that at a cutoff value of 30 mg/dL, CRP had 96% specificity and 90% sensitivity for detecting SBP with AUC= 0.91.

On the other hand, using a cut off value \geq 46.0 (µg/mL) for serum calprotectin in detecting SBP in our study revealed that serum calprotectin had a highly significant diagnostic performance with AUC 0.976 (P <0.001), sensitivity 100% and specificity 92%.

Rizk et al. ⁽²⁶⁾ demonstrated that at a cut off value of 270 mg/dl, ascitic fluid calprotectin had 86% specificity and 97.5% sensitivity for detecting SBP with AUC= 0.924. Whereas *Lutz et al.* ⁽²⁴⁾ found that the ratio of ascitic calprotectin to ascitic total protein with a cut-off value of 5.24 achieved a sensitivity of 90% and specificity of 81% in detecting SBP with AUC= 0.92 (p < 0.001).

On comparing the AUC of ROC curve of serum CRP and serum calprotectin in differentiating between group A (with SBP) and group B (with sterile cirrhotic ascites), it was found that P value was non-significant (P=0.242).

The previous findings indicate that serum calprotectin in combination with serum CRP correlate well and reliably with ascitic PMNL count in patients with SBP. Indeed, using serum calprotectin together with serum CRP is superior to the using of one of them alone and may serve as a surrogate marker for ascitic PMNL count and could be amenable to routine SBP screening.

CONCLUSION

Serum calprotectin in combination with serum CRP correlated well with ascitic PMNL count in patients with SBP. Using both markers together is superior to the using of one of them alone and may be useful for non-invasive diagnosis of SBP.

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