



Diagnostic Utility of Cell Block Method versus Cytospin Method in Pleural and Peritoneal Fluid Cytology

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Abstract

Background: Cytological evaluation of body cavity fluid is diagnostically challenging. Improved ethanol formalin fixative is used which offer excellent cytomorphological features. Cell blocks prepared from residual tissue fluids or effusion obtained by aspiration, can be useful adjunct to smear for establishing a more definitive cytopathologic diagnosis. .

Methods: A total of 170 fluid specimens were examined for cytospin smear and cell block method. Out of 170 fluids, 102 were peritoneal and 68 were pleural. Each fluid specimen was subjected to cytospin smear (CSS) technique, and 10% alcohol-formalin cell block (CB) technique. Overall morphological details, cellularity, architecture, nuclear and cytoplasmic details were studied in both CSS and CB techniques.

Results: In this study, analysis body fluid specimens using cytospin smear and cell block methods revealed that there is no difference between cytospin smear method and CB in defining the benign, fungal and inflammatory conditions. However, CB method could able to identity papillary pattern more efficiently than the cytospin method.

Conclusion: Although there was no statistical difference between the results obtained by the cytospin and cell block methods, cell block method in our study accurately diagnosed the cases which were missed or incompletely diagnosed on cytospin smear method. Thus cell block proved to be superior method for the study of effusion as compared to cytospin smear. As the cell blocks permit longer storage and additional analysis such as immunohistochemistry (IHC) and microarray, it should be adopted additionally for effusion cytology.

Keywords: Cell block, Cytospin, Cytodiagnosis, adenocarcinoma, Effusion.

Introduction

Effusion cytology is the study of individual cells from aspirated material for the diseases diagnoses, accurate diagnosis of cells of serous fluids is a major challenge and distinguishing benign from malignant may require meticulous screening^(1,2). Cytology gives the first indication of malignancy in one third of malignant effusions and due to bland morphological details of cells, overcrowding or overlapping of cells, cell loss, and processing methods of the laboratory, it is very difficult to achieve⁽¹⁾. The cytological examination of serous effusions is well-accepted as it provides a definitive diagnosis that helps in staging, prognosis and management of the patients in malignancies^(3,4). Further, it gives information about various inflammatory and non-inflammatory lesions of serous membrane⁽⁵⁻⁷⁾. Recent new biological discoveries and analytical breakthroughs led the usage of several nonstandard body fluids including saliva, peritoneal, pleural, synovial, wound, drain and washout fluids for the diagnosis of human diseases⁽⁸⁾. The information provided by body fluid analysis helps the clinician in formulating therapy and prognosis. There are a wide range of cytological techniques available to analyse body fluids, from simple direct smears, cytopsin smear to cell block methods. Selection of a particular method will depends on the aspiration preparation skill, location of the aspiration to the preparation lab and expertise of the cytopathologist⁽⁹⁾.

Cytopsin smear method is designed to concentrate cells that are found in small numbers. This method allows the cells to be spun at various speeds and times to ensure formation of a monolayer of cells for the best assessment of the cells⁽⁶⁾. The cell block (CB) technique is one of the oldest methods for the evaluation of body cavity fluids, in which small tissue fragments in a fluid specimen are processed to form a paraffin block^(2,5,6,10). The main advantages of the CB technique are preservation of tissue architecture and obtaining multiple sections for special stains and immunohistochemistry^(5,6,10-12). Further, a good

CB can be very useful for molecular diagnostic studies such as fluorescence in situ hybridization, polymerase chain reaction, and cDNA microarray analyses⁽¹³⁾. The present study is aimed to compare the relative usefulness of cytomorphological features by using cytopsin smear and cell block methods of body fluid analysis in the diagnosis of peritoneal and pleural effusions.

Materials and Methods

The present study was an observational prospective study conducted on 170 body fluid specimen collected by paracentesis for the diagnosis of effusion cytology by CSS and CB method that referred Histopathology and Cytology laboratory, Department of Pathology, Pt. J.N.M. Medical College & Dr. B.R.A.M. Hospital, Raipur (Chhattisgarh), India, between February 2014 and August 2015. Institutional ethics committee of Pt. J.N.M. Medical College, Raipur has approved this study. Informed written consent was collected from the relatives of the patients. Complete demographic information and a thorough medical history with relevant clinical details were collected for each sample. About 20 ml of fresh pleural/peritoneal fluid specimens were aspirated freshly from patients by following all aseptic precautions and local anesthesia. The gross examination of the fluid is done by describing the color, clarity, granularity, coagulum etc. Of the 20 ml specimen, only 3 ml was used for cytopsin smear method and rest was used for cell block preparation. The protocol adopted respectively for the cytopsin smear and cell block techniques were briefly as follows.

The cytopsin technique involves the use of cytopsin slides that are assembled with a slide filter card and sample delivery chamber, secured by a plastic clip. About 3ml sample was centrifuged at 700 rpm for 6 minutes in a Thermo Shandon cytocentrifuge. The cell suspension that spun onto a microscope slide was absorbed onto filter paper while the centrifuge is spinning. Smear were prepared & stained with Papanicolaou stain/ Hematoxylin and Eosin⁽⁵⁾. In cell block

technique, 10ml sample of fluid sample was mixed with equal volume of AF fixative (10 ml of 10% alcohol-formalin (i.e., nine parts of 90% alcohol and one part of 7.5 % formalin), kept for one hour and centrifuged at 2500 rpm for 15 minutes for obtain cell sediment. The supernatant was discarded and a further 3 ml of fresh 10% alcohol-formalin was added once again to the sediment, than kept for one day. On the following day, the sediment containing the cell button of the fluid sample was scooped out on to the Whatman's filter paper and processed in automatic tissue processor for routine histopathology section. From the paraffin embedded cell button (cell block), 4–6 μ thickness sections were prepared and stained with the Harris hematoxylin and eosin stain ⁽⁵⁾. The fluid specimens were categorized based on the morphological criteria including cellularity, arrangement of cells, nuclear and cytoplasmic details of each specimen. Final diagnosis of Benign, suspicious for malignancy and malignant effusion of the patient was made based on the clinical history, laboratory tests, radiological examination and cytological examination based on the cytospin and cell block techniques. To assess the difference between cytospin and cell block techniques chi-square test statistics were adopted.

Results

All the 170 body fluid specimens were subjected to the Cytospin smear and the CB techniques. Out of 170 patients, 67 patients (39.4%) were males and 103 patients (60.6%) were females with male:female ratio of 1:1.5. Of the 170 specimens analysed, 102 (60%) were of ascitic fluid and 68 (40%) were of pleural fluid (Table 1). Of all the effusions, 76.5% were turbid and 23.5% were clear (Table 1). Variation in the microscopic

impression obtained by cytospin smears and cell block methods in different fluid types was presented in table 2. There was no difference between cytospin smear method and CB in defining the benign, fungal and inflammatory conditions. However, CB method was more efficient (22.9%) in diagnosing malignant condition when compared to CSS method (20.0%). Further inquiry revealed that the both methods defined more neoplastic impression in ascitic fluids and nonneoplastic impression in pleural fluids (Table 2). Diagnostic characterization of neoplastic and non-neoplastic lesions using cytospin smear and cell block methods showed that there is no difference between these two methods for non-neoplastic conditions (Table 3). Variations in patterns of malignant cases diagnosed using cytospin smear and cell block methods was documented in table 4. Although both cytospin and CB methods are similar in defining the cellular pattern, CB method could able to identify papillary pattern more efficiently in the present sample (Table 4). The differences in the photomicrograph obtained through the cytospin smear and cell block is depicted in figure 1. Upon Haematoxylin and Eosin stain, cytospin smear showed cluster & scattered malignant cells, cells arranged in acinar pattern with nuclei were eccentrically pushed and mucin filled cytoplasm in hemorrhagic background giving the impression of adenocarcinoma of ovary. The same sample using cell block method and Haematoxylin and Eosin stain showed a hyperchromatic papillary cluster of malignant cells with stratification of cells and fibrovascular core with stratification of hyperchromatic nuclei in haemorrhagic background indicating the papillary adenocarcinoma of ovary (Figure 1).

Table 1: Characteristics of the fluid specimen used in the study

	Turbid fluid	Clear fluid	Total
Pleural fluid	50 (38.5)	18 (45.0)	68 (40.0)
Ascites fluid	80 (61.5)	22 (55.0)	102 (60.0)
Total	130	40	170

Table 2: Variation of microscopic impression obtained by cytopsin smears and cell block methods in different fluid types.

Microscopic impression	Cytospin smear method			Cell block method		
	Total	Pleural fluid	Ascites fluid	Total	Pleural fluid	Ascites fluid
Benign	45 (26.5)	14 (20.6)	31 (30.4)	45 (26.5)	14 (20.6)	31 (30.4)
Fungal	2 (1.2)	1 (1.5)	1 (1.0)	2 (1.2)	1 (1.5)	1 (1.0)
Inflammatory	62 (36.5)	34 (50.0)	28 (27.5)	62 (36.5)	34 (50.0)	28 (27.5)
Malignant	34 (20.0)	10 (14.7)	24 (23.5)	39 (22.9)	12 (17.6)	27 (26.5)
Suspicious of malignant	10 (5.9)	4 (5.9)	6 (5.9)	5 (2.9)	2 (2.9)	3 (2.9)
Others	17 (10.0)	5 (7.4)	12 (11.8)	17 (10.0)	5 (7.4)	12 (11.8)

Table 3: Diagnostic characterization of neoplastic and non-neoplastic lesions using cytopsin smear and cell block methods.

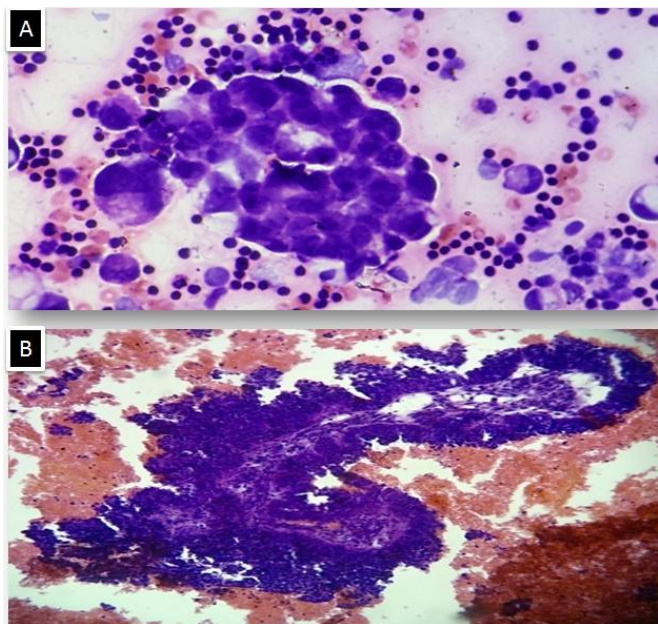
Diagnosis	Cytospin	Cell block
Non-neoplastic		
RMs	18 (16.5)	18 (16.5)
RMs with inflammation	27 (24.8)	27 (24.8)
Acute inflammation	3 (2.8)	3 (2.8)
Chronic inflammation	40 (36.7)	40 (36.7)
Acute & chronic inflammation	19 (17.4)	19 (17.4)
Fungal	2 (1.8)	2 (1.8)
Chi square p value		1.00
Neoplastic		
ADCA	25 (56.8)	28 (63.6)
SCC	4 (9.1)	5 (11.4)
RCT	3 (6.8)	3 (6.8)
MM	1 (2.3)	1 (2.3)
PDCA	1 (2.3)	2 (4.5)
STBM	10 (22.7)	5 (11.4)
Chi square p value		0.492

RMs: Reactive mesothelial cells; ADCA: Adenocarcinoma; SCC: Squamous cell carcinoma; RCT: Round cell tumor; MM: malignant melanoma; PDCA: Poorly differentiated carcinoma; STBM: Suspected to be malignant.

Table 4: Variations in patterns of malignant cases diagnosed using cytopsin smear and cell block methods.

S.No.	Subtypes of malignancy	Various pattern showed	Cytospin	Cell block
1.	Adenocarcinoma	Scattered/clustered/acinar	17 (50)	16 (41)
		Papillary & acinar pattern	4 (11.8)	4 (10.3)
		Mucinous papillary	0 (0)	2 (5.1)
		Serous papillary with psammoma bodies	0 (0)	1 (2.6)
		Signet ring carcinoma	2 (5.9)	2 (5.1)
		Brochioloalveolar	0 (0)	1 (2.6)
		Poorly differentiated adenocarcinoma	2 (5.9)	2 (5.1)
2.	Squamous cell carcinoma	Moderately differentiated	2 (5.9)	2 (5.1)
		Poorly differentiated	2 (5.9)	3 (7.7)
3.	Malignant round cell tumor		3 (8.8)	3 (7.7)
4.	Malignant melanoma		1 (2.9)	1 (2.6)
5.	Poorly differentiated carcinoma		1 (2.9)	2 (5.1)

Figure 1: Photomicrograph of cytopsin smear and cell block methods.



Cytopsin smear indicating adenocarcinoma of ovary (H & E - 400x); B. Cell block indicating the papillary adenocarcinoma of ovary (H & E - 400x).

Discussion

Analysis of 170 body fluid specimens using cytopsin smear and cell block methods revealed that there is no difference between cytopsin smear method and CB in defining the benign, fungal and inflammatory conditions. However, CB method could able to identify papillary pattern more efficiently than the cytopsin method. Further, photomicrograph obtained through the cell block method provided better impression of malignancy than that depicted by cytopsin smear method.

Multiple independent studies conducted on effusion cytology have shown that the cytopsin and cellblocks methods are superior to conventional method in diagnosing the effusions. Conventional smears failed in making conclusive diagnosis due to lack of morphological details of the representative cells in the sample⁽⁶⁾. Whereas in cytopsin preparations allow the preservation of cellular details and reduce the overlapping of cells⁽¹⁴⁾. Manifestation of glandular formations and acinar groupings in cell block method made it as a superior to smear based methods^(5,15). Direct

comparison of effusion analysis by cytopsin and cell block methods revealed that there is no difference between these methods⁽¹⁶⁾. Scope for performing immunohistochemistry and microarray on cellblocks is an added advantage of cellblock method⁽¹⁷⁾. Although, cell block preparations facilitate better diagnosis of lesions, sometimes fails in providing conformation and lead to suspicion of malignancy. In the present study, five cases (2.94%) could not achieve final diagnosis by all available clinical details & morphological features. A comparative study reported that 0.67% cases failed to achieve final diagnosis by all modalities⁽¹⁰⁾. Adenocarcinoma is the commonest malignancy found in body effusions^(1,6,10,18-20). The present study report well appreciated acinar, mucinous papillary, serous papillary, bronchioloalveolar, signet ring carcinoma patterns by cell block than cytopsin smears. Among hemorrhagic malignant effusions, cell blocks give better appreciation of malignant cells due to formation of two layers in sediment. But care should be taken during embedding the cell button that cutting surface should be the upper part of the sediment to avoid more hemorrhagic background, or glacial acetic acid could be mixed in hemorrhagic effusions to obtain clear cellular details.

Conclusions

Although there was no statistical difference between the results obtained by the cytopsin and cell block methods, cell block method in our study accurately diagnosed the cases which were missed or incompletely diagnosed on cytopsin smear method and recognition of specific histological patterns of diseases is possible by using cell block method. It is also useful for special stains and immunohistochemistry and can give morphological details by preserving the architectural patterns. Storage of slides & blocks for retrospective studies is easy by using cell block method. Thus cell block proved to be superior method for the study of effusion as compared to cytopsin smear.

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